

GENETIC AND MOLECULAR DETERMINANTS OF PEMPHIGUS VULGARIS AND THEIR THERAPEUTIC IMPLICATIONS

Marius IRIMIE¹

Abstract: *Pemphigus vulgaris (PV) is a rare, potentially life-threatening autoimmune blistering disease driven by pathogenic IgG autoantibodies targeting desmosomal cadherins (primarily desmoglein-3 and desmoglein-1), leading to loss of keratinocyte adhesion (acantholysis). Beyond antibody-mediated mechanisms, convergent evidence supports a strong genetic contribution to disease susceptibility, phenotype, and therapeutic response. This expanded narrative review synthesizes classical and emerging genetic determinants of PV including HLA class II associations, non-HLA immune modifiers, desmosomal/epithelial susceptibility genes, genome-wide association study (GWAS) loci, epigenetic and transcriptomic signals and integrates contemporary therapeutic advances (rituximab, FcRn inhibitors, BTK inhibition, and antigen-specific cell therapies). We highlight genotype-phenotype relationships, ethnic differences in risk alleles, translational biomarkers, and future directions for precision medicine in PV.*

Key words: *pemphigus vulgaris, immunogenetics, HLA, desmoglein, ST18, FcRn inhibitors, rituximab*

1. Introduction

Pemphigus vulgaris (PV) is the most common form of pemphigus and is characterized by mucosal and/or cutaneous blistering that rapidly evolves into painful erosions. Prior to immunosuppressive therapy, pemphigus was frequently fatal; contemporary treatment has reduced mortality but at the cost of significant morbidity from chronic disease, infections, and long-term corticosteroid exposure. The epidemiology of PV exhibits striking geographic and

ethnic variation, with consistently higher incidence in certain populations (e.g., individuals of Jewish, Mediterranean, Middle Eastern, South Asian, and some East Asian ancestries), supporting a strong inherited component [1-3]. In recent years, an increase in the number of cases of pemphigus vulgaris has been observed in Romania, with personal observations indicating a higher prevalence in ethnic groups of Asian origin.

The canonical model of PV pathogenesis centers on circulating IgG autoantibodies against desmogleins. However, several

¹ Faculty of Medicine, *Transilvania University of Braşov*

observations cannot be fully explained by humoral autoimmunity alone: (i) variability in age of onset and clinical phenotype, (ii) differences in autoantibody specificity and subclass distribution, (iii) heterogeneity in relapse risk and response to rituximab, and (iv) population-specific allele associations. Over the last two decades, immunogenetic and genomic studies have provided a richer, polygenic view of PV that implicates antigen presentation, immune checkpoints, cytokine networks, B-cell biology, and keratinocyte-intrinsic susceptibility pathways [4-6].

2. Clinical Spectrum, Diagnosis, and Outcome Measures

PV may present as mucosal-dominant disease (often oral erosions preceding skin lesions), cutaneous-dominant disease, or mucocutaneous disease. The desmoglein compensation hypothesis provides a biologic explanation for mucosal predilection: DSG3 is highly expressed in the basal and suprabasal layers of mucosa, whereas DSG1 is more abundant in superficial epidermis. Accordingly, patients with predominant anti-DSG3 reactivity may exhibit mucosal-dominant disease, while additional anti-DSG1 reactivity is associated with cutaneous involvement [2, 3].

Diagnosis relies on clinicopathologic correlation. Histopathology typically reveals suprabasal acantholysis (“row of tombstones”) and direct immunofluorescence demonstrates intercellular IgG/C3 deposition in the epithelium. Serologic assays (ELISA for anti-DSG1/3) support diagnosis and longitudinal monitoring. Standardized outcome measures such as the Pemphigus

Disease Area Index (PDAI) facilitate assessment of severity and response across clinical trials and real-world cohorts [2].

3. Immunopathogenesis: beyond autoantibodies

PV is driven by pathogenic IgG autoantibodies, but multiple immune pathways converge to sustain autoimmunity. Antigen presentation by HLA class II molecules activates autoreactive CD4⁺ T cells that provide B-cell help via costimulation and cytokine support, promoting class switching and differentiation into antibody-secreting cells. Autoantibody binding to desmogleins triggers signaling cascades (p38 MAPK, Src-family kinases, EGFR pathways) and cytoskeletal reorganization, resulting in desmosome disassembly and keratinocyte detachment (figure 1) [7, 8].

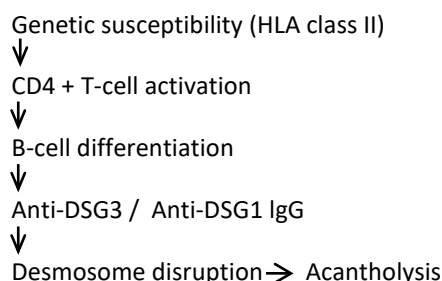


Fig. 1. Immunopathogenic cascade in pemphigus vulgaris (PV)

Recent mechanistic work reinforces that keratinocytes are not passive targets: they respond to immune injury by altering adhesion, stress-response, and inflammatory gene expression programs. This provides a rationale for therapies targeting signaling pathways (e.g., EGFR and downstream kinases) and for integrating epithelial biology into genetic models of PV [7, 8].

4. HLA Class II Susceptibility and Genotype-Phenotype Links

The most robust genetic associations in PV reside in the HLA class II locus on chromosome 6p21. Across diverse populations, HLA-DRB1*04:02 and HLA-DQB1*05:03 are repeatedly associated with increased risk. These alleles are thought to present immunodominant desmoglein peptides to CD4⁺ T cells with high efficiency, lowering the threshold for loss of tolerance. Importantly, HLA risk alleles vary in frequency across ancestries, which shapes both susceptibility and the

landscape of autoimmune specificity [9]. Genotype-phenotype correlations have increasingly been documented: DRB1*04:02 has been associated with higher anti-DSG3 levels and mucosal-only disease, while DQB1*05:03 may correlate with anti-DSG1 reactivity and higher rates of mucocutaneous disease. Emerging data further identify population-specific signals such as DRB1*08:04 in patients of African ancestry, emphasizing the need for inclusive cohorts and careful interpretation of linkage disequilibrium (table 1) [9].

Selected HLA alleles associated with PV across populations (representative examples) Table 1

HLA allele	Populations frequently reported	Typical phenotype association (reported)	Notes
DRB1*04:02	Ashkenazi Jewish; non-Jewish Caucasian; Hispanic; others	Higher anti-DSG3; mucosal-dominant disease	Strongest and most replicated PV risk allele
DQB1*05:03	South Asian; East Asian; many non-Jewish cohorts	Higher anti-DSG1; mucocutaneous disease	Often co-occurs with specific DRB1 alleles
DRB1*14:01 / 14:04	Mediterranean; Middle Eastern; Asian cohorts	Susceptibility signal; variable phenotype	May represent multiple related risk haplotypes
DRB1*08:04	Patients of African ancestry (reported in US cohorts)	Elevated anti-DSG3; phenotype variable	Highlights underrepresented genetic architecture

5. Non-HLA Immune Genetic Modifiers

Candidate-gene studies and GWAS-informed analyses implicate non-HLA immune genes as modifiers of PV risk and disease expression. Variants in cytokine genes (e.g., TNF, IL6, IL10) may influence inflammatory tone, B-cell differentiation, and persistence of antibody-secreting cells. Polymorphisms in immune checkpoint and costimulatory pathways (e.g., CTLA4, ICOS, CD40/CD40L) may

modulate T-cell activation thresholds and germinal-center responses. Fc gamma receptor variants (FCGR2A/FCGR3A) can alter immune-complex handling and effector functions [10]. A recurring limitation is population size: PV is rare, and many association studies have modest cohorts and heterogeneous ancestry, yielding variable replication. Nevertheless, convergent biology supports a polygenic model in which HLA risk enables pathogenic antigen presentation

while a background of immune regulatory variation shapes magnitude, chronicity, and treatment responsiveness of the autoantibody response [10].

6. Non-HLA Immune Genetic Modifiers

Genome-wide Association Studies (GWAS) have broadened PV genetics beyond classical immune loci. A prominent and repeatedly discussed locus is ST18 (suppression of tumorigenicity 18), encoding a transcription factor implicated in inflammation and apoptosis. Risk variants near ST18 have been associated with increased expression in keratinocytes, potentially amplifying cytokine responses and sensitizing epithelium to antibody-mediated injury. Population-specific association studies continue to explore ST18 polymorphisms and their functional impact [11].

Other GWAS-linked or fine-mapped signals implicate genes in interferon and antigen-presentation pathways (e.g., STAT4, IRF8, TAP2), pointing to cross-talk between innate cues and adaptive autoimmunity. Integrating GWAS with transcriptomics and epigenetic datasets is increasingly viewed as necessary to prioritize causal genes and mechanisms [11].

7. Desmosomal Genes, Barrier Biology, and Epitope Spreading

Although PV is defined by autoimmunity against desmosomal proteins, genetic variation within epithelial adhesion molecules may influence antigenicity and mechanical resilience. Polymorphisms in DSG3/DSG1 or in desmosomal partners (desmocollins, plakophilins) could affect expression, conformation, or exposure of epitopes, thereby shaping susceptibility to

autoantibody binding or promoting tissue damage under stress. Such variants may also facilitate epitope spreading broadening the autoimmune response to additional junctional targets over time [12].

This epithelial dimension aligns with experimental data showing that keratinocyte signaling and stress pathways strongly modulate acantholysis. Targeting keratinocyte-intrinsic pathways (e.g., EGFR signaling) has shown promise in ex vivo human skin models, supporting the concept of combining immunomodulation with barrier-stabilizing strategies [7].

8. Epigenetics, Transcriptomics, and Systems-level Signals

Epigenetic dysregulation may contribute to PV by altering immune cell differentiation programs and keratinocyte inflammatory responses. Studies have reported aberrant DNA methylation patterns and histone modifications in peripheral blood mononuclear cells, although causal relationships remain to be established. Transcriptomic profiling of blood and lesional tissues has identified immune activation signatures and potential transcriptional “hot spots,” suggesting that gene-expression changes could help distinguish PV from other autoimmune diseases and may yield biomarkers for disease activity or relapse risk [5].

Future work integrating genomics (GWAS/WES/WGS), epigenomics, proteomics, and single-cell immune profiling is expected to refine PV endotypes molecularly defined subgroups that may respond differently to B-cell depletion, FcRn blockade, or targeted pathway inhibition [13, 14].

9. Therapeutic Implications and Precision Medicine

Rituximab (anti-CD20) has transformed PV management and is widely recommended as first-line therapy for moderate-to-severe disease in contemporary guidelines (table 2, figure 2). Long-term follow-up studies of rituximab-based first-line regimens suggest durable remission with reduced reliance on prolonged corticosteroids in many patients. However, relapses still occur and may reflect incomplete depletion of autoreactive memory B cells and long-lived plasma cells, or rapid

immune reconstitution in genetically predisposed individuals [15-20].

FcRn inhibitors (e.g., efgartigimod) represent a mechanistically distinct approach that accelerates IgG clearance and can rapidly reduce pathogenic anti-DSG titers. Early-phase clinical trials in pemphigus demonstrate rapid disease control and improvements in PDAI alongside reductions in total IgG and anti-desmoglein autoantibodies. Because FcRn blockade does not directly target B-cell production, combination or sequencing strategies with B-cell-directed agents are being explored [15, 16], [21, 22].

Table 2
Emerging and established targeted therapeutic strategies in PV (mechanism-oriented overview)

Strategy	Examples	Primary target	Rationale in PV	Evidence type (recent)
B-cell depletion	Rituximab; other anti-CD20	CD20+ B cells	Reduce autoantibody generation	Guidelines; RCTs; long-term follow-up; real-world
IgG recycling blockade	Efgartigimod (Fc fragment); other FcRn inhibitors	FcRn	Rapidly lower pathogenic IgG	Phase II trial; translational studies
BCR signaling inhibition	Rilzabrutinib	BTK	Suppress B-cell activation & innate signaling	Phase II data; program updates
Antigen-specific B-cell elimination	DSG3-CAAR T cells	Autoreactive anti-DSG3 B cells	Precision removal of pathogenic clones	Early-phase clinical trial
Keratinocyte pathway modulation	EGFR pathway inhibitors (experimental)	EGFR/kinase signaling	Reduce acantholysis signaling cascades	Ex vivo / translational studies

Small-molecule pathway inhibitors (e.g., BTK inhibition with rilzabrutinib) aim to suppress B-cell receptor signaling and innate immune activation while potentially reducing corticosteroid

exposure. Early phase results showed clinical activity and acceptable safety, although trial outcomes have been mixed across programs [23]. Additional targeted strategies include inhibition of CD40-

CD40L interactions, blockade of inflammatory kinases, and approaches to modulate keratinocyte signaling [8].

Antigen-specific cellular therapies, including DSG3 chimeric autoantibody receptor (CAAR) T cells, represent a precision concept: selectively eliminating

autoreactive B cells while sparing protective humoral immunity. Ongoing early-phase trials are evaluating feasibility and safety in mucosal-dominant PV [24–26]. If successful, such strategies could offer durable, drug-free remission in carefully selected patients.

Therapeutic evolution in PV (conceptual)

1950s–: systemic corticosteroids

1980s–2000s: conventional steroid-sparing agents (AZA/MMF/CYC)

2010s: rituximab (anti-CD20) → first-line for moderate–severe PV

2020s: targeted agents (FcRn inhibitors, BTK inhibitors, CAAR-T, pathway inhibitors)

Fig.2. Conceptual timeline of therapeutic milestones in pemphigus vulgaris (PV).

10. Limitations and Future Directions

Despite major progress, several gaps persist. Genetic studies remain uneven across global populations, and many cohorts are underpowered for fine-mapping or multi-omic integration. Heterogeneity in clinical phenotype definitions (mucosal vs mucocutaneous) and treatment exposure can obscure genotype–phenotype relationships. Future studies should prioritize: (i) multinational consortia with harmonized phenotyping, (ii) ancestry-aware fine-mapping and functional validation, (iii) single-cell immune profiling of blood and lesional tissues before and after targeted therapies, and (iv) development of predictive biomarkers integrating genetics, autoantibody kinetics, and immune reconstitution dynamics.

From a translational perspective, a realistic near-term goal is to define clinically useful endotypes: for example,

patients with rapid relapse after rituximab versus those with sustained remission; patients with predominant mucosal disease and high anti-DSG3 titers versus those with broader antigen spreading. Such endotypes could guide early combination therapy (e.g., rituximab + FcRn inhibitor), maintenance strategies, and monitoring intensity.

References

1. Kridin K, Schmidt E. Epidemiology of pemphigus. *JID Innov.* 2021 Feb 20; 1(1):100004. doi: 10.1016/j.xjidi.2021.100004.
2. Schmidt E, Kasperkiewicz M, Joly P. Pemphigus. *Lancet.* 2019 Sep 7; 394(10201):882–894. doi: 10.1016/S0140-6736(19)31778-7.
3. Kasperkiewicz M, Ellebrecht CT, Takahashi H, et al. Pemphigus. *Nat Rev Dis Primers.* 2017 May 11;3:17026. doi: 10.1038/nrdp.2017.26.

4. Vodo D, Sprecher E. The genetic basis of pemphigus vulgaris. *JEADV Clin Pract.* 2023; 2:203-212. <https://doi.org/10.1002/jvc2.129>
5. Baker J, Seiffert-Sinha K, Sinha AA. Patient genetics shape the autoimmune response in the blistering skin disease pemphigus vulgaris. *Front Immunol.* 2023; 13:1064073. <https://doi.org/10.3389/fimmu.2022.1064073>.
6. Russu EA, Popa LG, Burcea A, et al. Genetic and molecular insights into pemphigus vulgaris with oral involvement: implications for biologic therapy. *Rom J Oral Rehabil.* 2025; 17(2):559-574. doi: 10.62610/RJOR.2025.2.17.51.
7. Egu DT, Schmitt T, Ernst N, et al. EGFR inhibition by erlotinib rescues desmosome assembly and ameliorates PV IgG-induced acantholysis. *J Invest Dermatol.* 2024 Nov; 144(11): 2440-2452. doi: 10.1016/j.jid.2024.03.040.
8. Payne AS, Vieira SM. Bruton's Tyrosine Kinase Inhibition in Pemphigus: An Embattled Frontier. *J Invest Dermatol.* 2024 Aug; 144(8): 1667-1670. doi: 10.1016/j.jid.2024.03.014.
9. Didona D, Maglie R, Eming R, Hertl M. Pemphigus: current and emerging treatment strategies. *Front Immunol.* 2019 Jun 25; 10:1418. doi: 10.3389/fimmu.2019.01418.
10. Gjølborg TT, Mester S, Calamera G, et al. Targeting the Neonatal Fc Receptor in Autoimmune Diseases: Pipeline and Progress. *BioDrugs.* 2025 May; 39(3):373-409. doi: 10.1007/s40259-025-00708-2.
11. Al-Hasnawi ZA, Al-Drobie B. Assessing ST18 gene polymorphisms (rs17315309, rs2304365) in Iraqi patients with pemphigus vulgaris. *J Med Life.* 2024; 17(1): 81-86. doi: 10.25122/jml-2023-0227.
12. Schmidt E, Zillikens D. Modern diagnosis of autoimmune blistering diseases. *Autoimmun Rev.* 2010 Dec; 10(2):84-9. doi: 10.1016/j.autrev.2010.08.007.
13. Ma G, Crowley AR, Heyndrickx L, et al. Differential effects of FcRn antagonists on the subcellular distribution of IgG and immune complexes. *JCI Insight.* 2024 May 7; 9(10):e176166. doi: 10.1172/jci.insight.176166
14. Yang CW, Xia T, Tan Q, et al. Evaluating the clinical impact of FcRn inhibition in IgG-mediated autoimmune disease. *Front Immunol.* 2025 Oct 2; 16:1656937. doi: 10.3389/fimmu.2025.1656937.
15. Tedbirt B, Tedbirt B, Maho-Vaillant M, et al. Sustained remission without corticosteroids among patients with pemphigus who had rituximab as first-line therapy: follow-up of the Ritux 3 trial. *JAMA Dermatol.* 2024; 160(3):290-296. doi: 10.1001/jamadermatol.2023.5679
16. Branisteanu DE, Lotti T, Colac Boțoc C, et al. Real-world outcomes with rituximab vs conventional therapy in pemphigus vulgaris. *Front Med.* 2025; 12: 1691897. doi: 10.3389/fmed.2025.1691897.
17. Szymanski K, Kowalewski C, Walecka I, et al. Eight years of follow-up of rituximab in pemphigus. *J Clin Med.* 2025 Oct 16; 14(20):7318. doi: 10.3390/jcm14207318.
18. Cao S, Yang B, Wang Z, et al. Efficacy, safety, and B-cell depletion capacity of ultralow-dose rituximab in

- pemphigus. *J Am Acad Dermatol*. 2025 Sep; 93(3):634-643. doi: 10.1016/j.jaad.2025.05.1374.
19. Joly P, Horwath B, Patsatsi A, et al. Updated S2K guidelines on the management of pemphigus vulgaris and foliaceus initiated by the European Academy of Dermatology and Venereology (EADV). *J Eur Acad Dermatol Venereol*. 2020; 34(9):1900-1913. <https://doi.org/10.1111/jdv.16752>
 20. Joly P, Maho-Vaillant M, Prost-Squarcioni C, et al. First-line rituximab combined with short-term prednisone versus prednisone alone for the treatment of pemphigus (Ritux 3): a prospective, multicentre, parallel-group, open-label randomised trial. *Lancet*. 2017 May 20;389(10083):2031-2040. doi: 10.1016/S0140-6736(17)30070-3.
 21. Gwathmey KG, Broome CM, Goebeler M, et al. Safety profile of efgartigimod from global clinical trials across multiple immunoglobulin G-mediated autoimmune diseases. *Expert Rev Clin Immunol*. 2025 May;21(5):627-638. doi: 10.1080/1744666X.2025.2497840.
 22. Goebeler M, Bata-Csörgő Z, De Simone C, et al. Treatment of pemphigus vulgaris and foliaceus with efgartigimod, a neonatal Fc receptor inhibitor: a phase II multicentre, open-label feasibility trial. *Br J Dermatol*. 2022; 186(3):429-439. doi:10.1111/bjd.20782
 23. Murrell DF, Caux F, Patsatsi A, et al. Efficacy and safety of rilzabrutinib in pemphigus. *J Invest Dermatol*. 2024 Aug; 144(8): 1762-1771.e6. doi: 10.1016/j.jid.2024.02.023.
 24. ClinicalTrials.gov. NCT04422912: DSG3-CAART (DesCAARTes™) trial in mucosal pemphigus vulgaris. <https://www.clinicaltrials.gov/study/NCT04422912>, Accessed December 28, 2025.
 25. Kale B, Khosravi-Maharlooei M, Perna F, et al. Shifting gears with CAR T cells for autoimmune diseases. *Mol Ther*. 2024 Feb 7; 32(2):261-263. doi: 10.1016/j.ymthe.2024.01.014.
 26. Rangel-Peláez C, Martínez-Gutiérrez L, Tristán-Manzano M, et al. CD19 CAR-T cell therapy: a new dawn for autoimmune rheumatic diseases? *Front Immunol*. 2024 Dec 17; 15:1502712. doi: 10.3389/fimmu.2024.1502712.